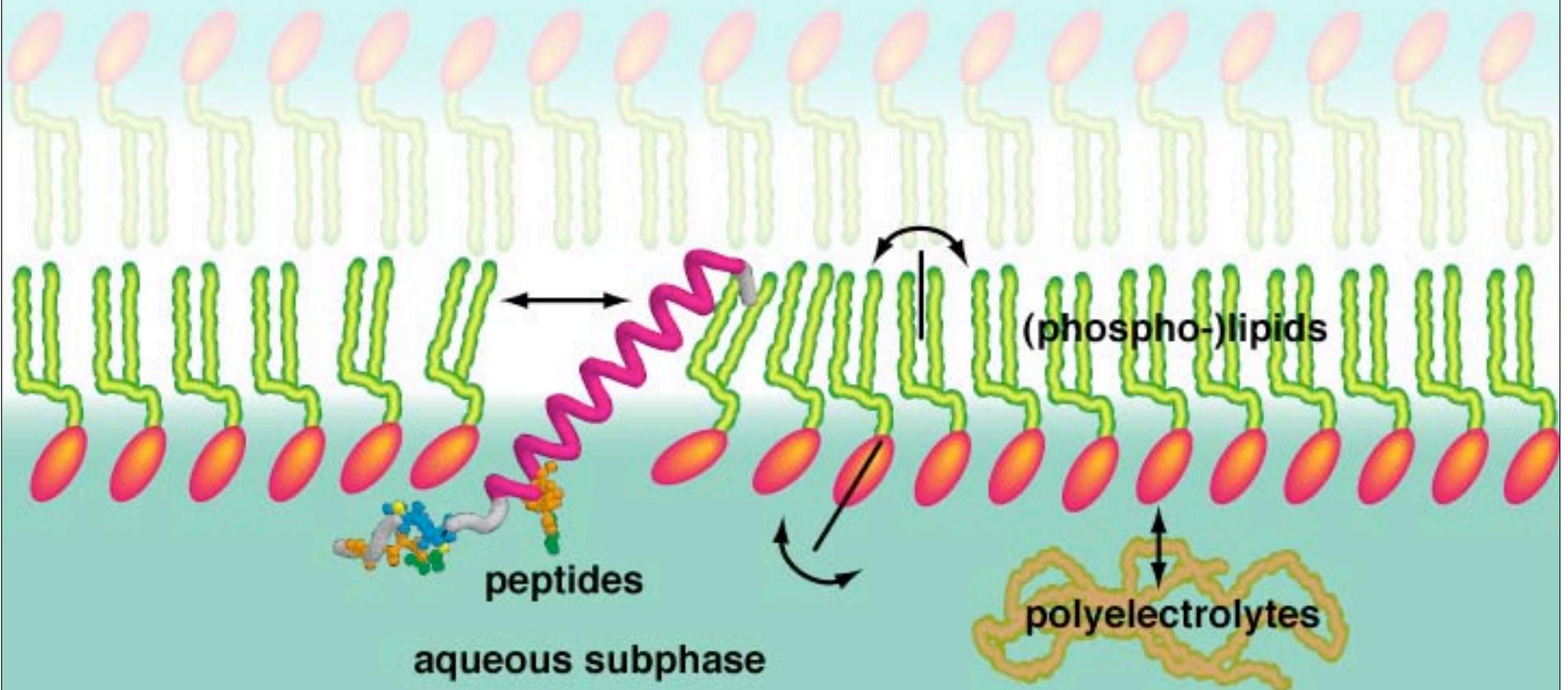


Surface-Sensitive Neutron Scattering as a Tool for the Molecular-Scale Characterization of Biomimetic Membranes



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& Physics Dept., Carnegie Mellon University, Pittsburgh, PA



*A Biotechnology Research Partnership
Funded by
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RR14812*

Joins high-performance computing with a unique
neutron diffractometer/reflectometer...

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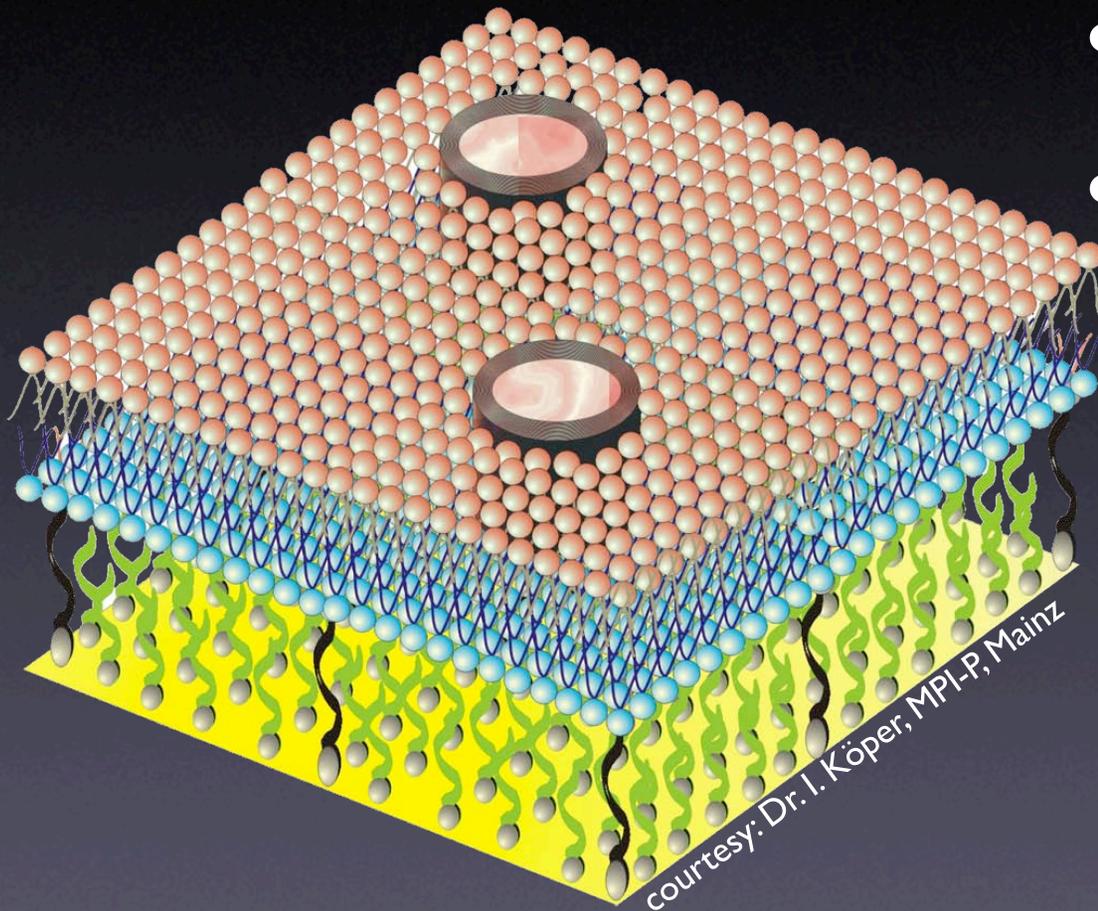
Carnegie Mellon University

Los Alamos National Laboratory

NIH (NIAAA)

... for structural studies of membrane-active proteins
in their native environment, including experiment-
validated studies of dynamics

Tethered Bilayer Lipid Membranes (tBLMs)

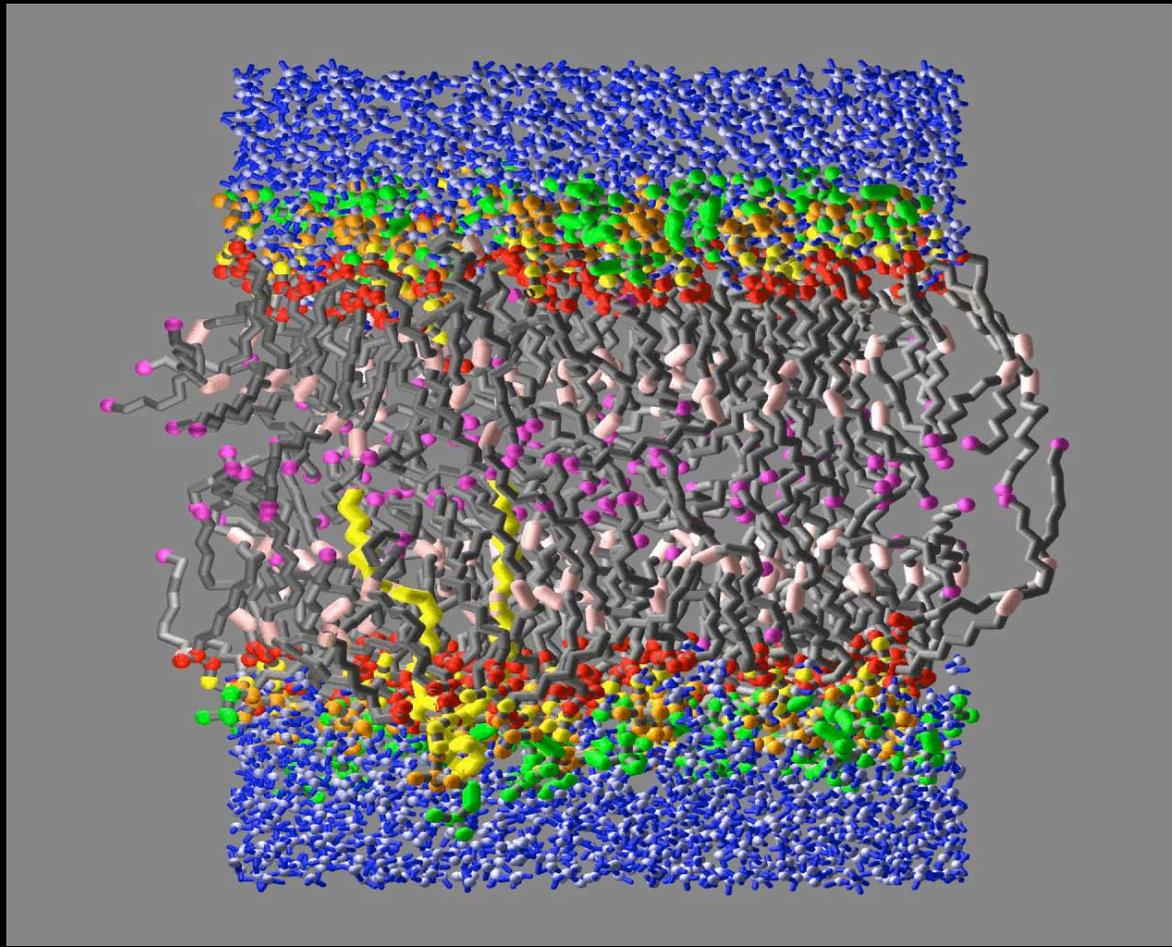


- comprehend biological processes
- utilize self-assembly for technical applications

→ are they indeed
what we *think* they are?

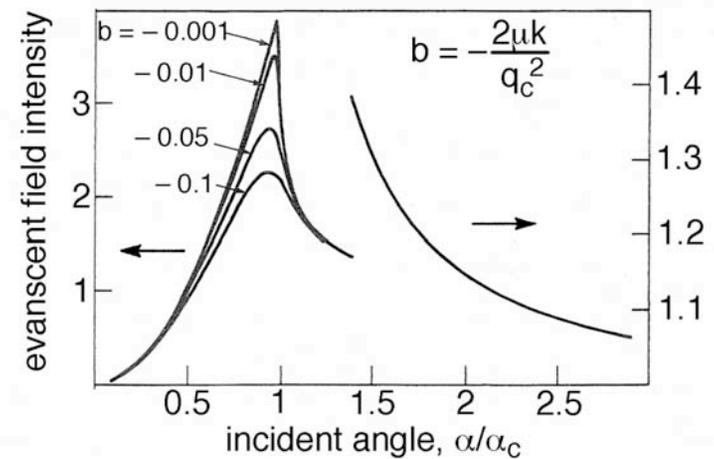
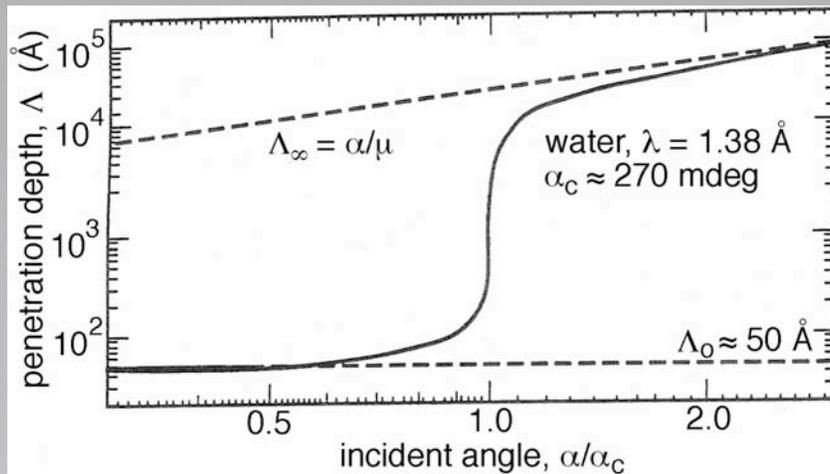
A Characteristic of Biomembrane Systems: High Thermal Disorder!

MD simulation: DOPC bilayer membrane

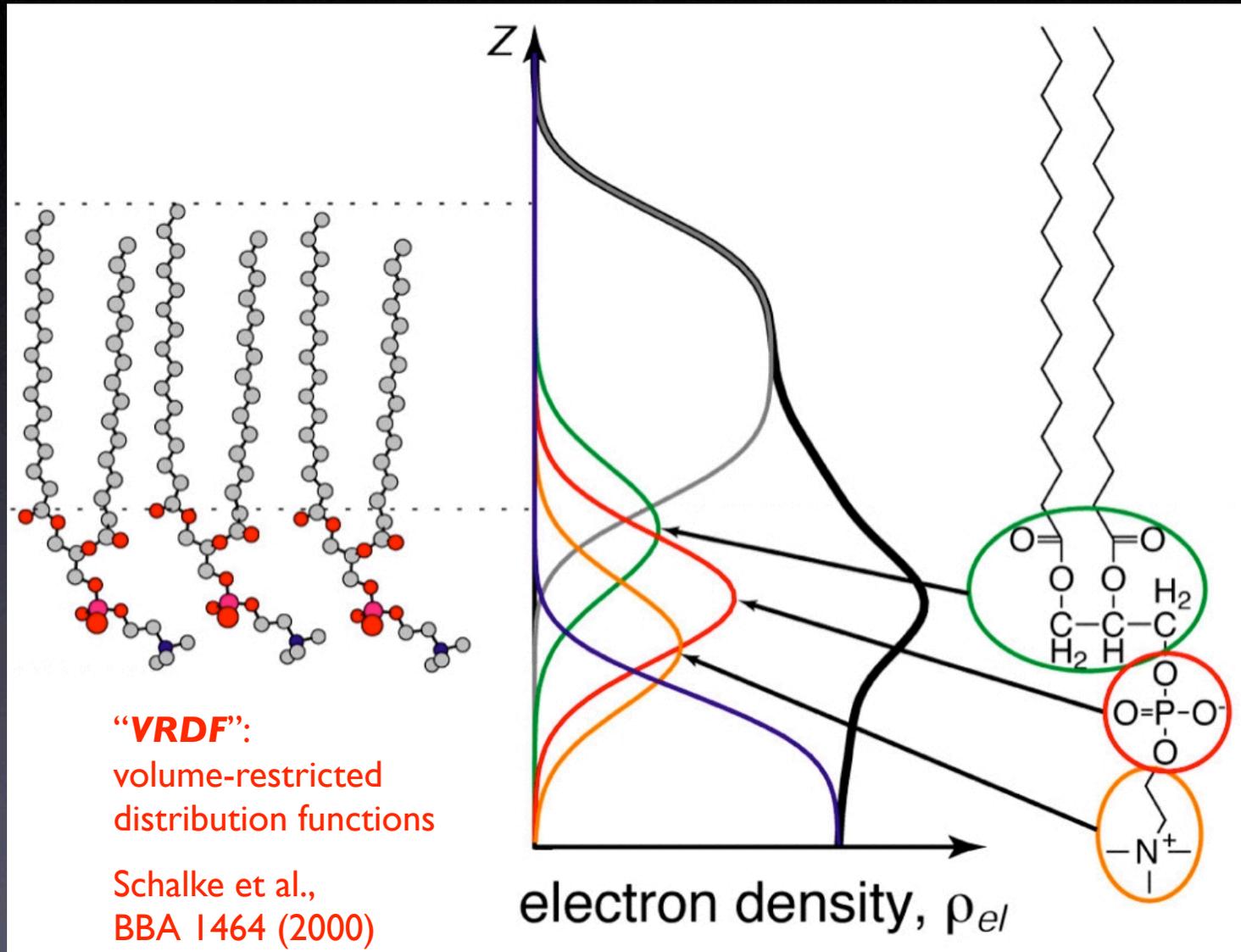


courtesy D J Tobias (UC Irvine)

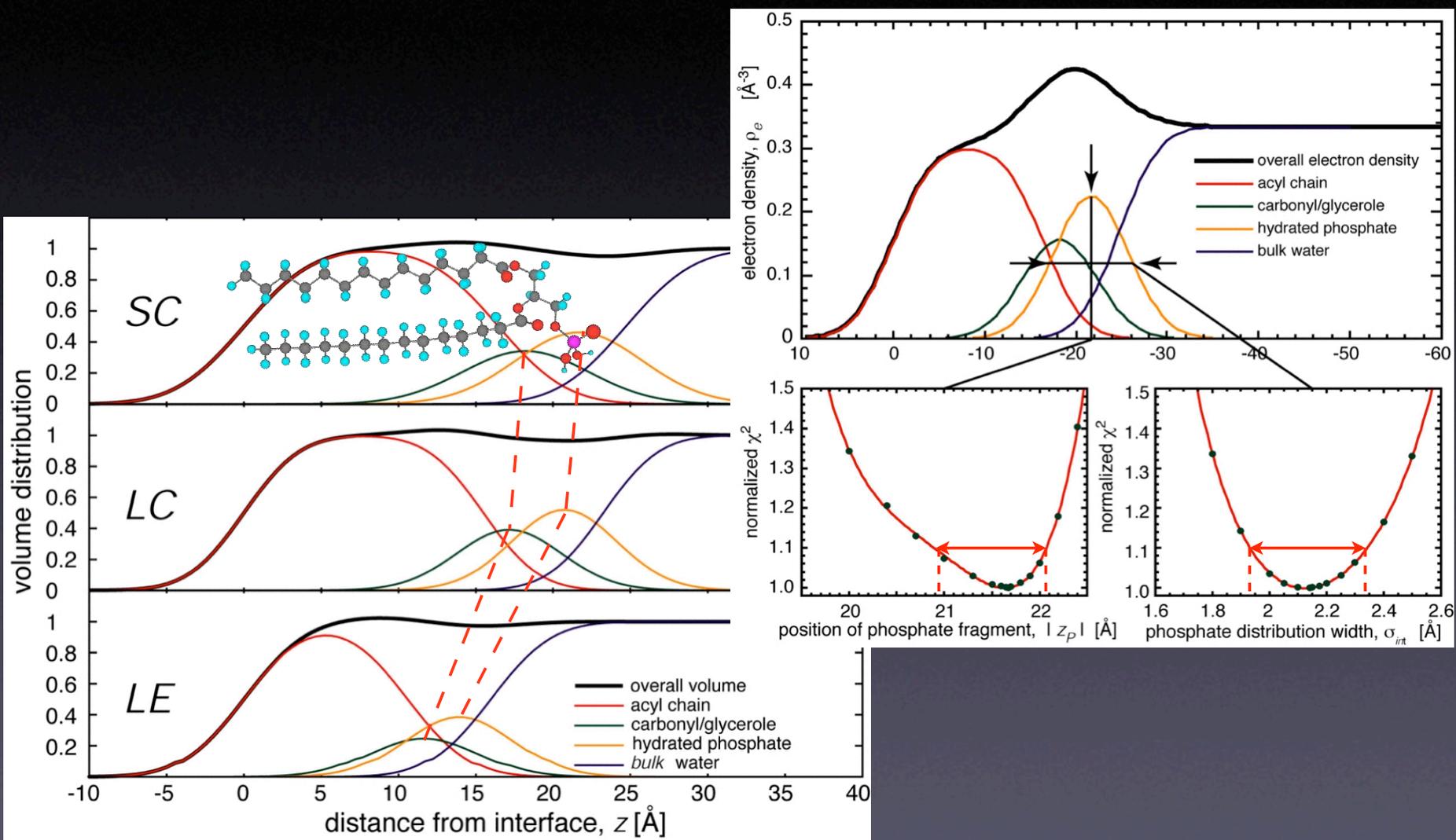
Surface-Sensitive Neutron and X-ray Scattering



Composition-Space Refinement Modelling

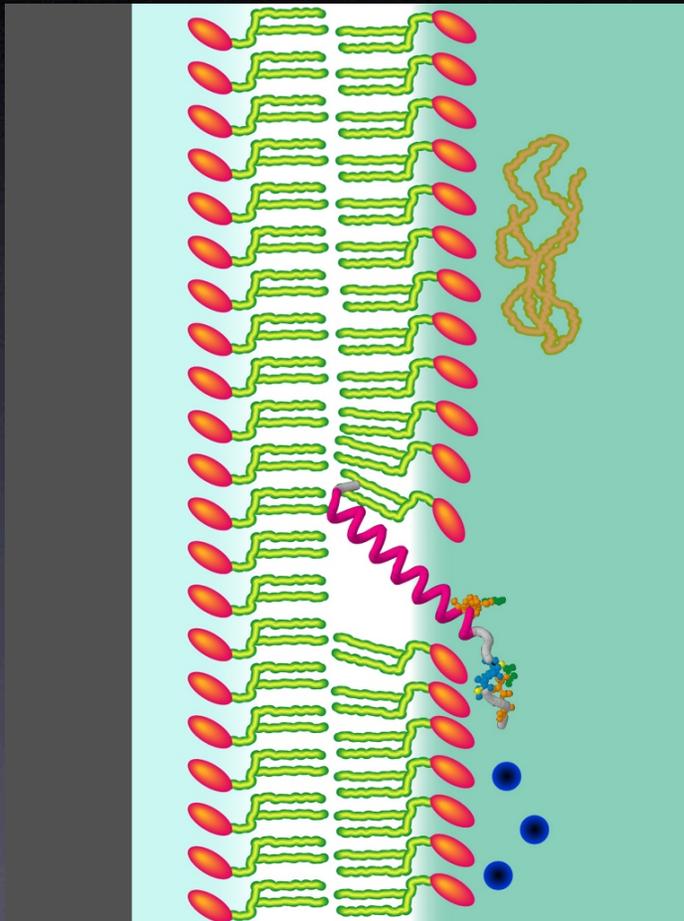


Model Performance and Credibility: DMPA



(x-ray characterization of Langmuir monolayers)

Solid-Supported Bilayer Membranes: Variations on a common theme

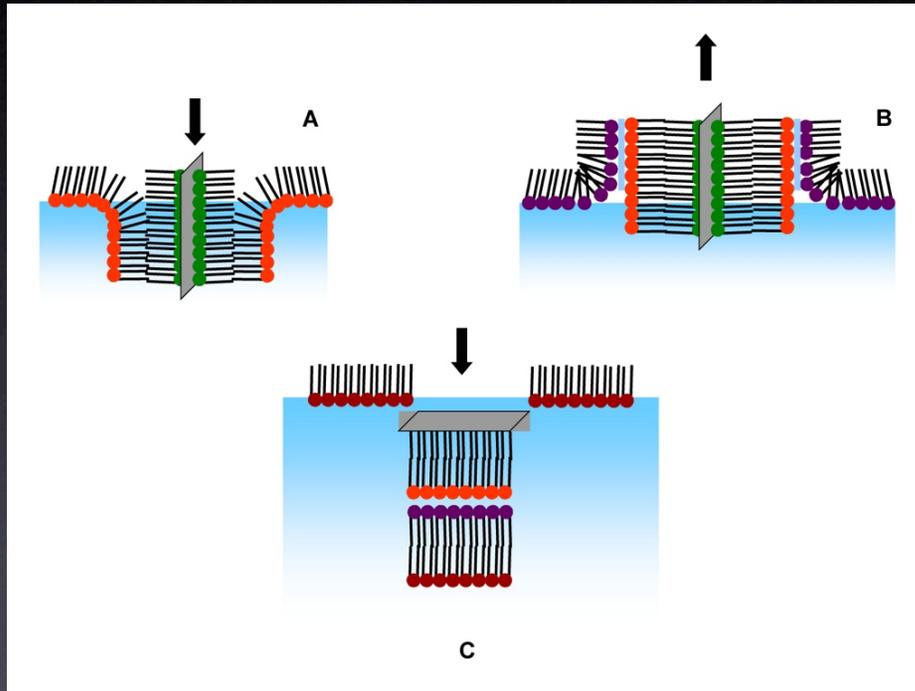


- *Adsorbed bilayers (Tamm & McConnell – and many others): vesicle fusion, etc.*
- *Floating bilayers (Roser; Fragneto & coworkers): LB transfer*
- *Tethered bilayers (Knoll, Ringsdorf, Cornell – and many others): thiol (gold), silane (quartz) chemistries; PEG spacers, etc.*

Neutron scattering:

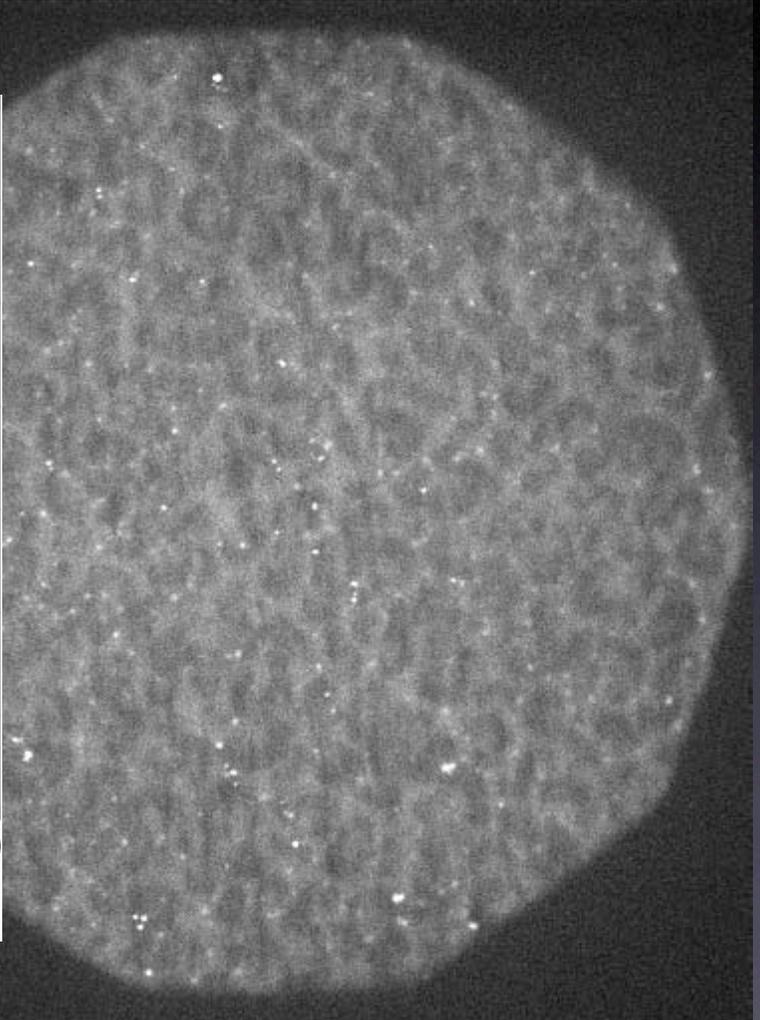
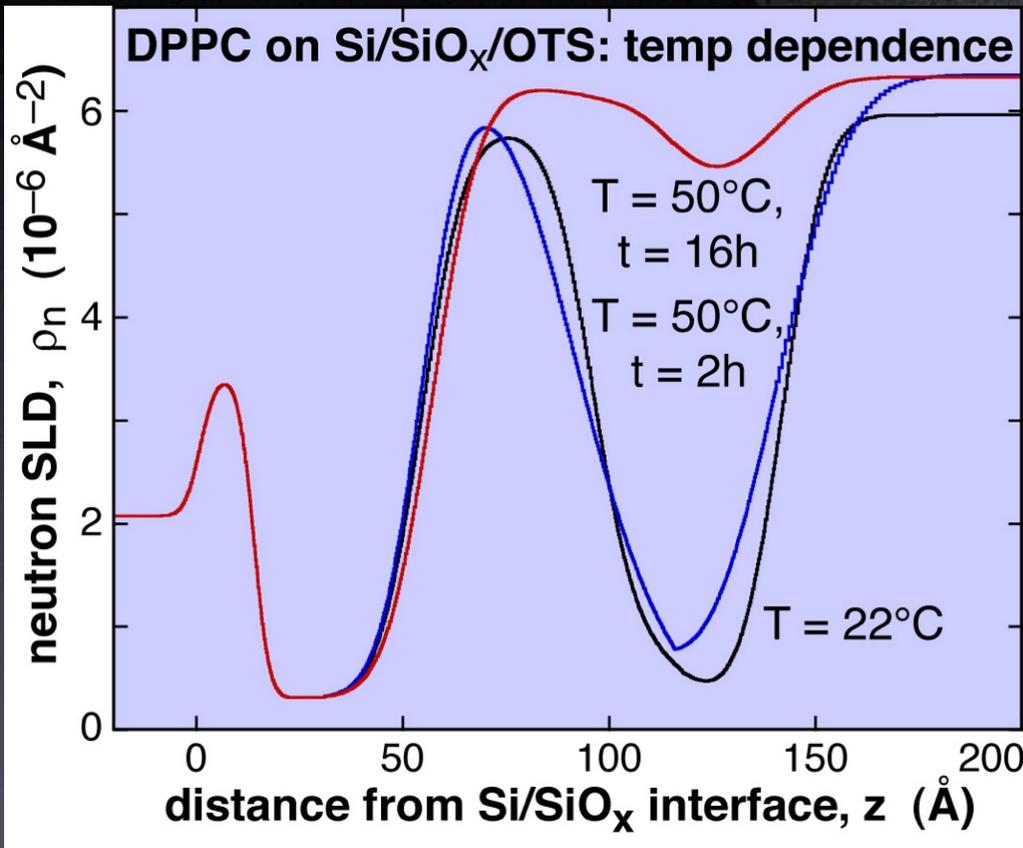
- *deeply penetrating*
- *non-destructive*
- *molecular sensitivity & resolution*

“Floating” Bilayers as Fluid Membrane Models



- *Step-by-step preparation (Langmuir trough)*
- *Retains undulation dynamics*
- *Transfer often incomplete*

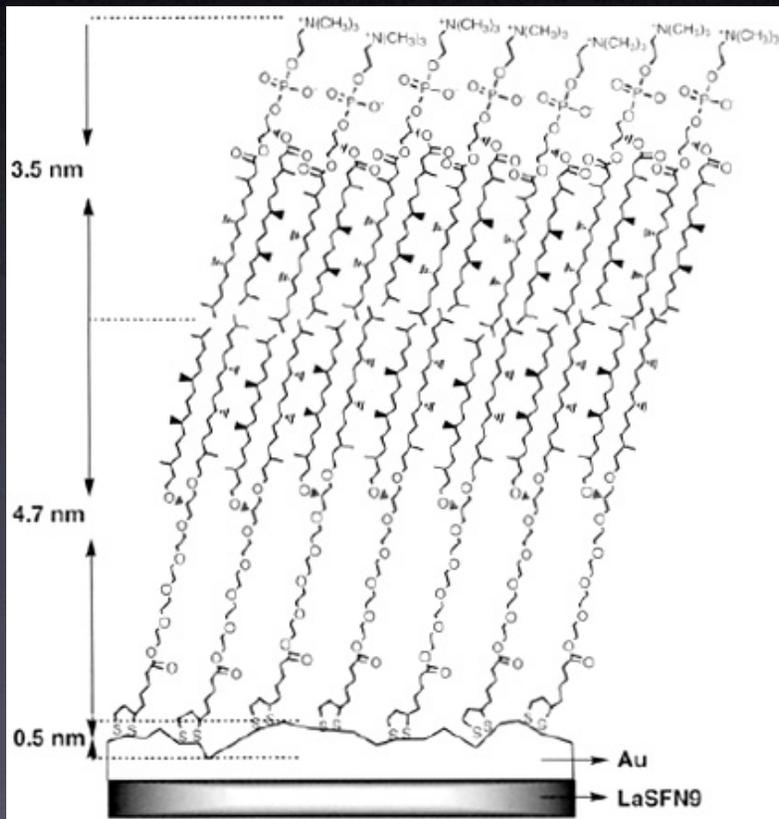
“Floating” Bilayers as Fluid Membrane Models



A. Kerth/D.J. McGillivray, unpublished

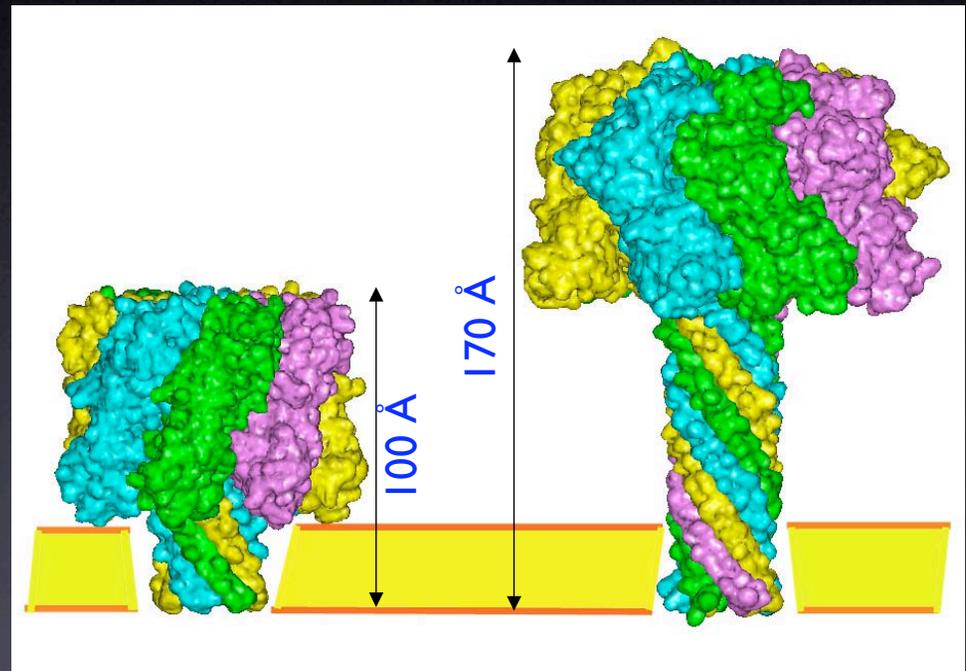
†BLMs and their Functionalization

Schiller et al., Angew. Chemie 42 (2003)



(collaboration with W. Knoll, J. Kasianowicz)

e.g., bacterial toxins:

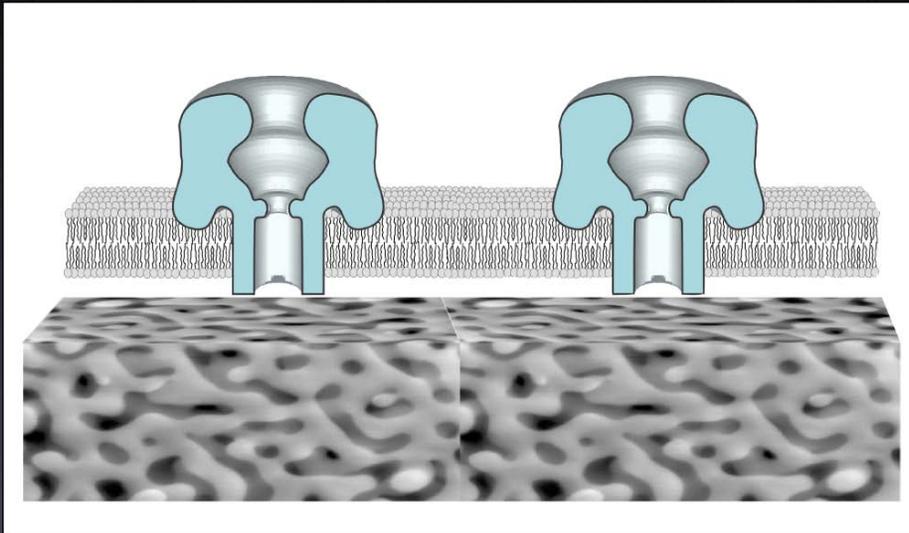


hemolysin
(Song et al.,
Science 1996)

B. anthracis PA63: putative
membrane association
(Nguyen, J Biomol
Struct Dyn 2004)

Bilaterally Accessible, Scaffolded *Fluid* Membranes

→ studies of signal transduction & amplification; biosensoric devices



active NSF-NIRT, together with

Mike Paulaitis (OSU ChemEng),
Jonah Erlebacher (JHU Mats Sci & Eng), and
John Kasianowicz (NIST Electron Elt Eng Lab)

scaffold structures:

porous gold, S-layer sheet crystals

goal:

incorporation into stabilized
membrane systems

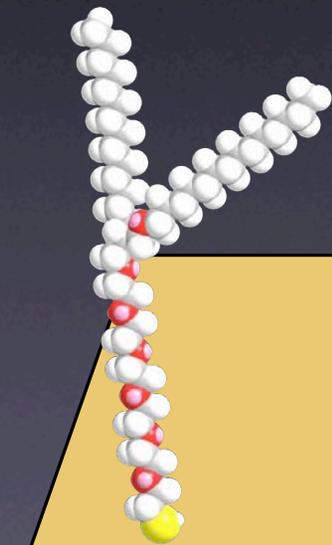
as a basis for

- molecular sensor systems
- pharmaceutical screening

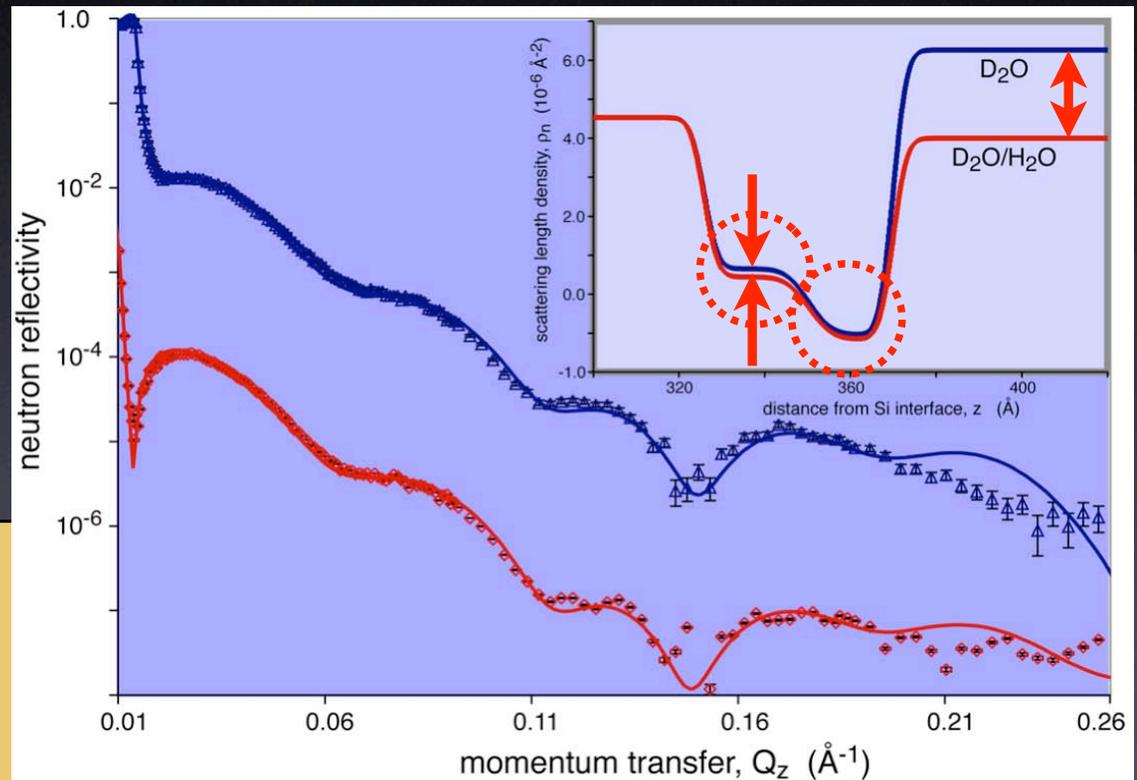
Tethered Bilayer Membranes (on continuous gold only, so far)

WC14:

A thiolated oligo-EG
dialkane for grafting on
Au surfaces (synthesis:
Dave Vanderah *et al.*)

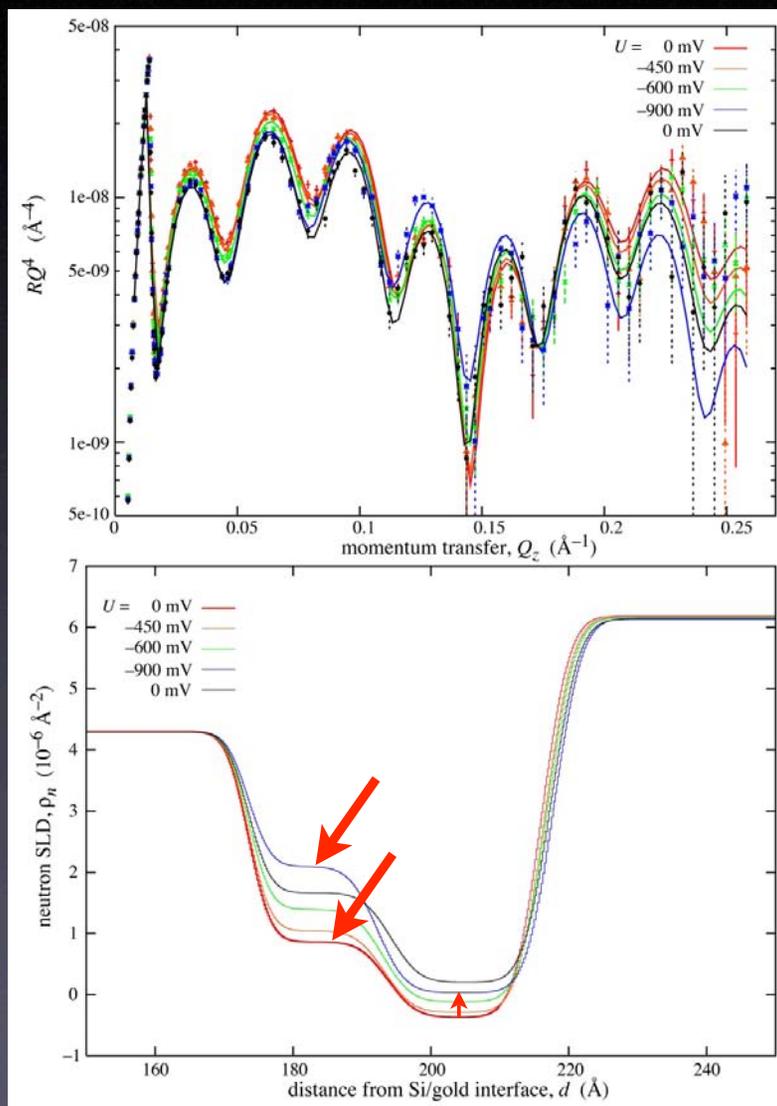


WC14 monolayer on Au (Si) under water



- extremely dense alkane packing
- $\ll 10$ vol% solvent in proximal space

Control of Hydration by Electric Field: Structural assessment as a function of electrostatic potential

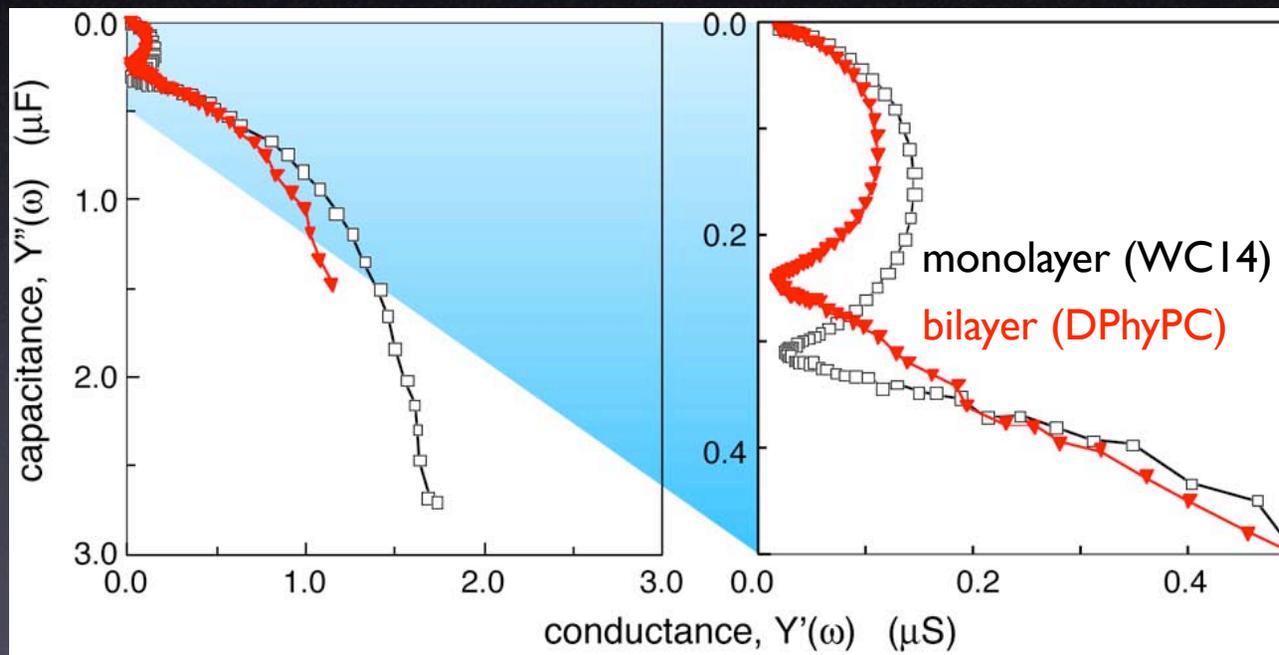


- DPTL monolayer under water
- “cleaning” of surface w/ positive pulse
- long-term stability at $\Phi \sim 4 \times 10^8 \text{ V/cm}$
- submembrane space significantly more hydrated upon return to $U = 0 \text{ V}$
- (but alkyl region more disordered: hydration through creation of defects!)

Köper & McGillivray, unpublished data

Bilayer Completion: “Rapid Solvent Exchange”

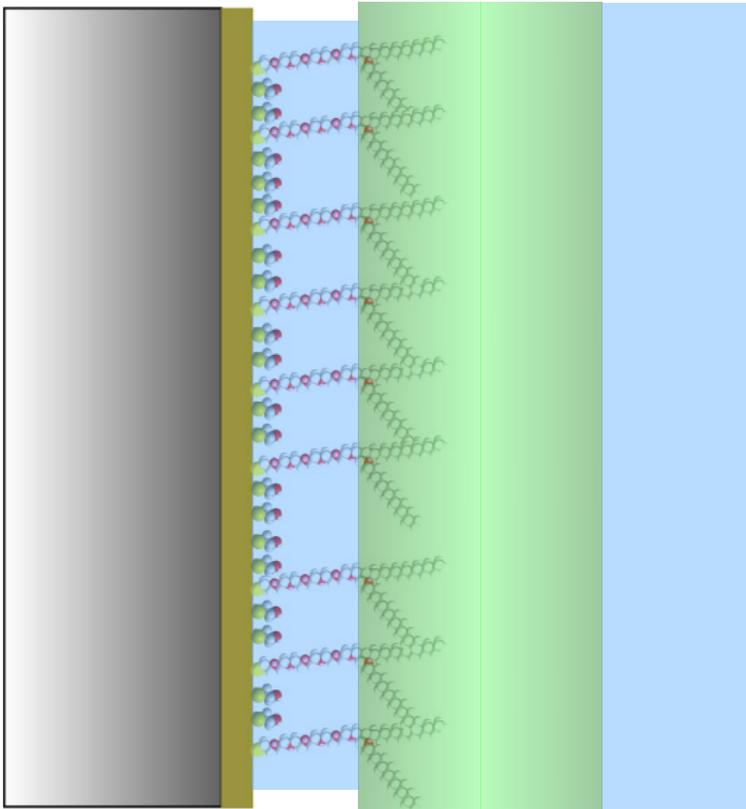
Nyquist diagrams of monolayer and bilayer



- capacitance ($0.75 \mu\text{F}/\text{cm}^2$) and conductance indicate bilayer formation
- EIS and NR: bilayer > 90% complete

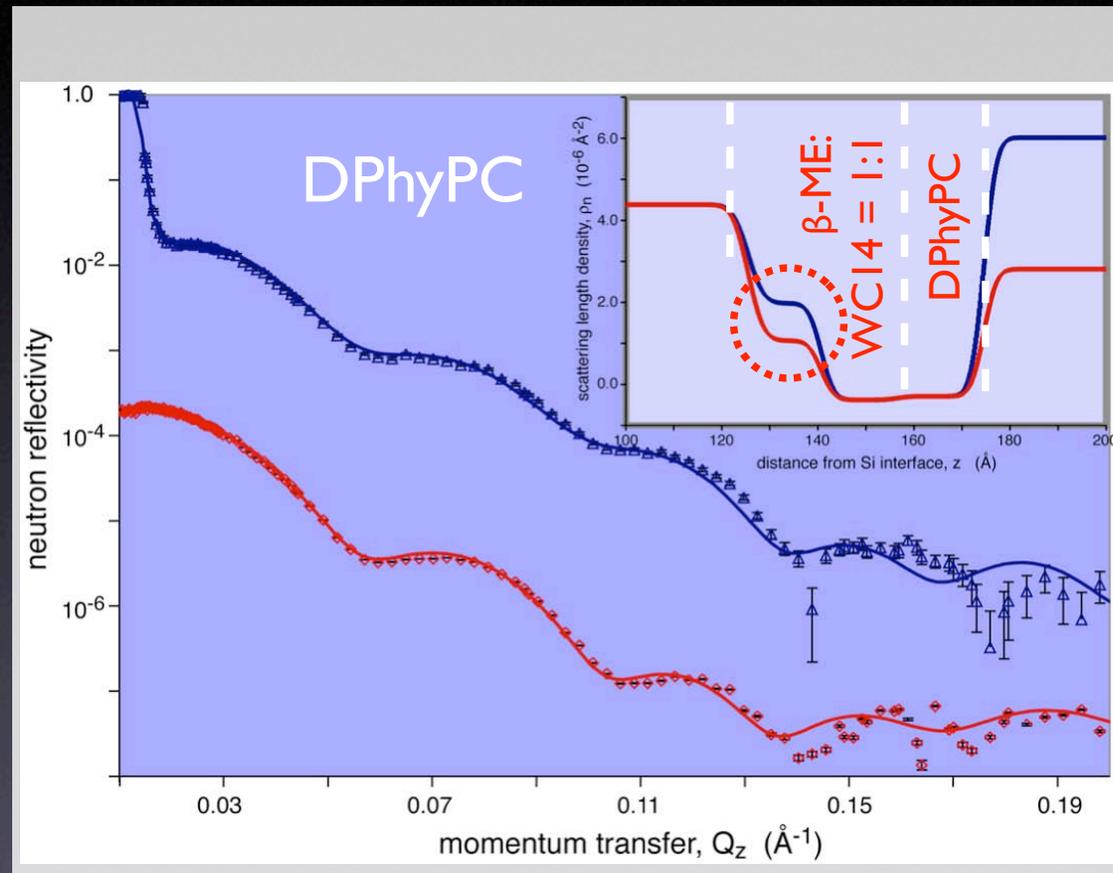
“rapid solvent exchange”:
(Cornell et al., *Nature* 387, 1997, 580)

Backfilling: β -Mercaptoethanol



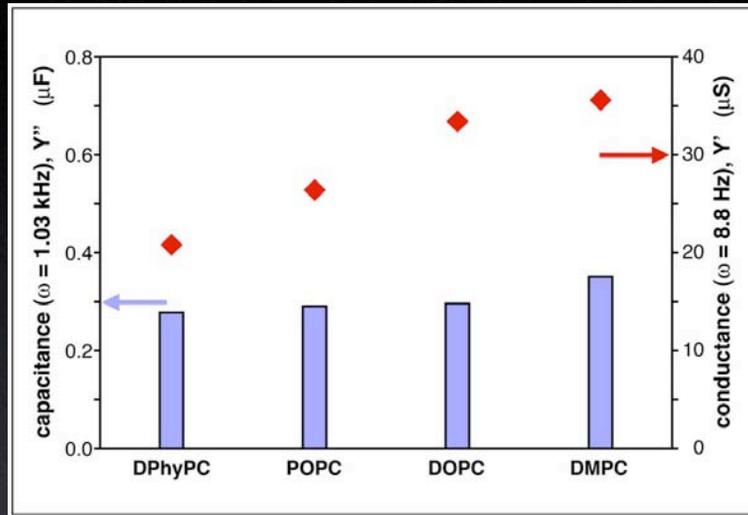
- does **not** work with vesicle fusion (low contact angle of the monolayer)
- works **well** with rapid solvent exchange

tBLM's Based on Mixed Monolayers of WCI4 and β -ME

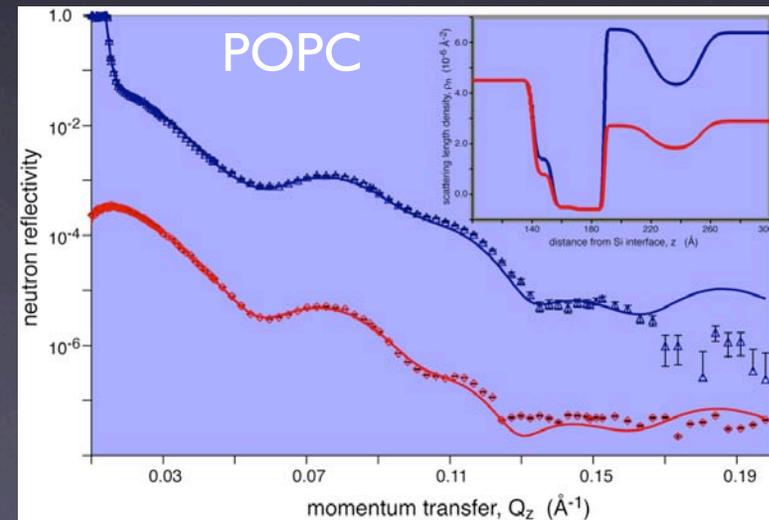
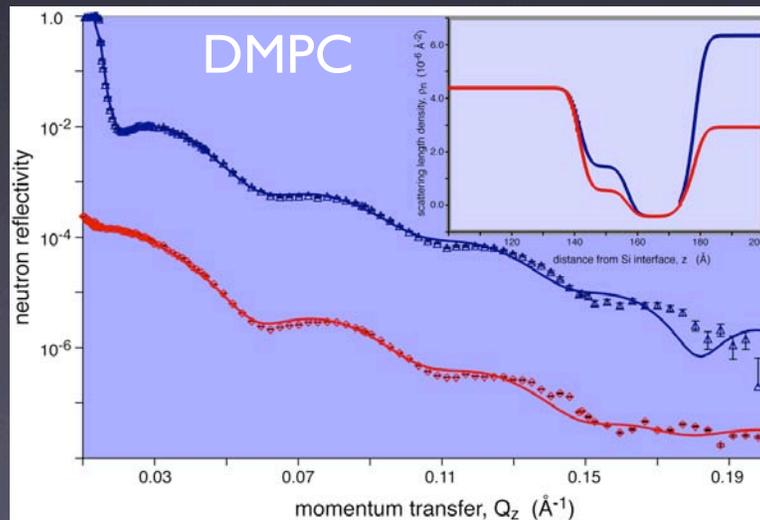


- Y', Y'' of mixed monolayer \gg pure WCI4
- Y', Y'' of bilayer (DPhyPC) based on mixed monolayer \sim as for pure WCI4
- β -ME introduces solvent in proximal space ($> 50\%$ solvent by vol.)

Dependence on Cover Lipid

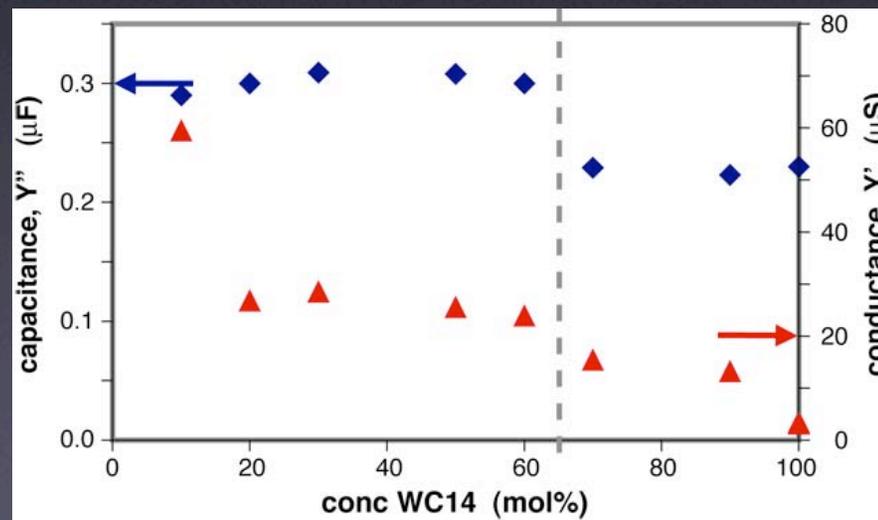
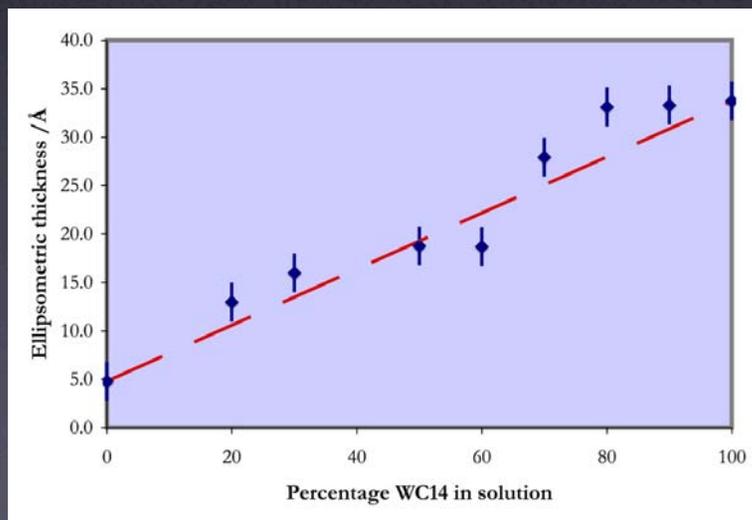
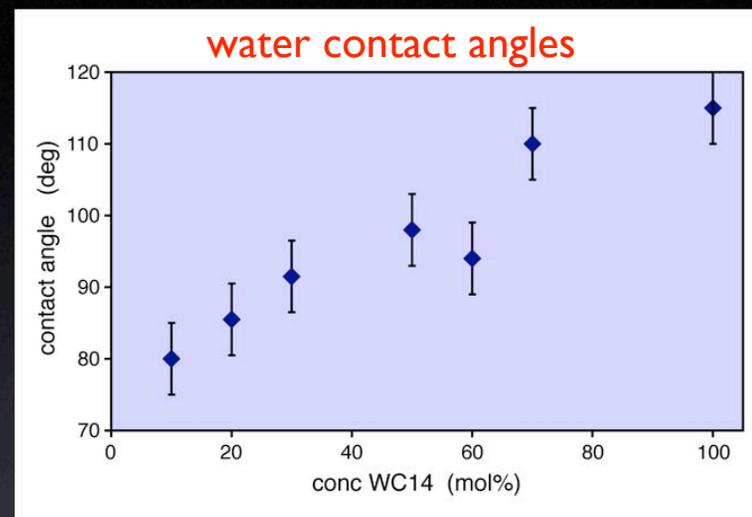


- DPhyPC, DMPC, POPC, DOPC all form complete bilayers on WC14/ β -ME (1:1)
- Y'' almost invariant for all PC's
- Y' depends significantly on lipid
- POPC forms partial overlayer

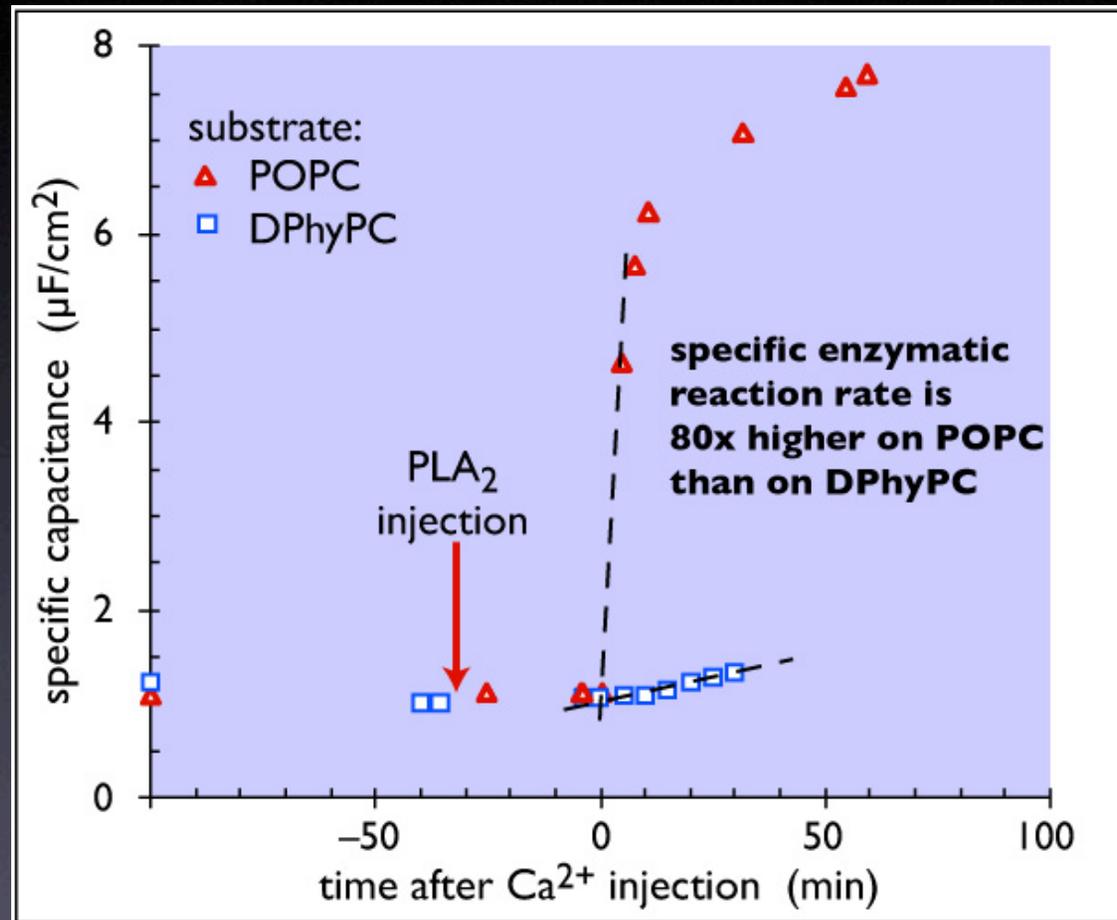


Influence of Backfiller Concentration

- *surface* concentration controlled by WC14: β -ME ratio in *deposition* solution
- significant amounts of WC14 on surface even at low solution conc
- partial fill-in of surface-tethered monolayer during solvent exchange
- structural/functional transition as a function of WC14 conc



Challenging with Phospholipase PLA₂: A Sensitive Amplifier of Local Defects

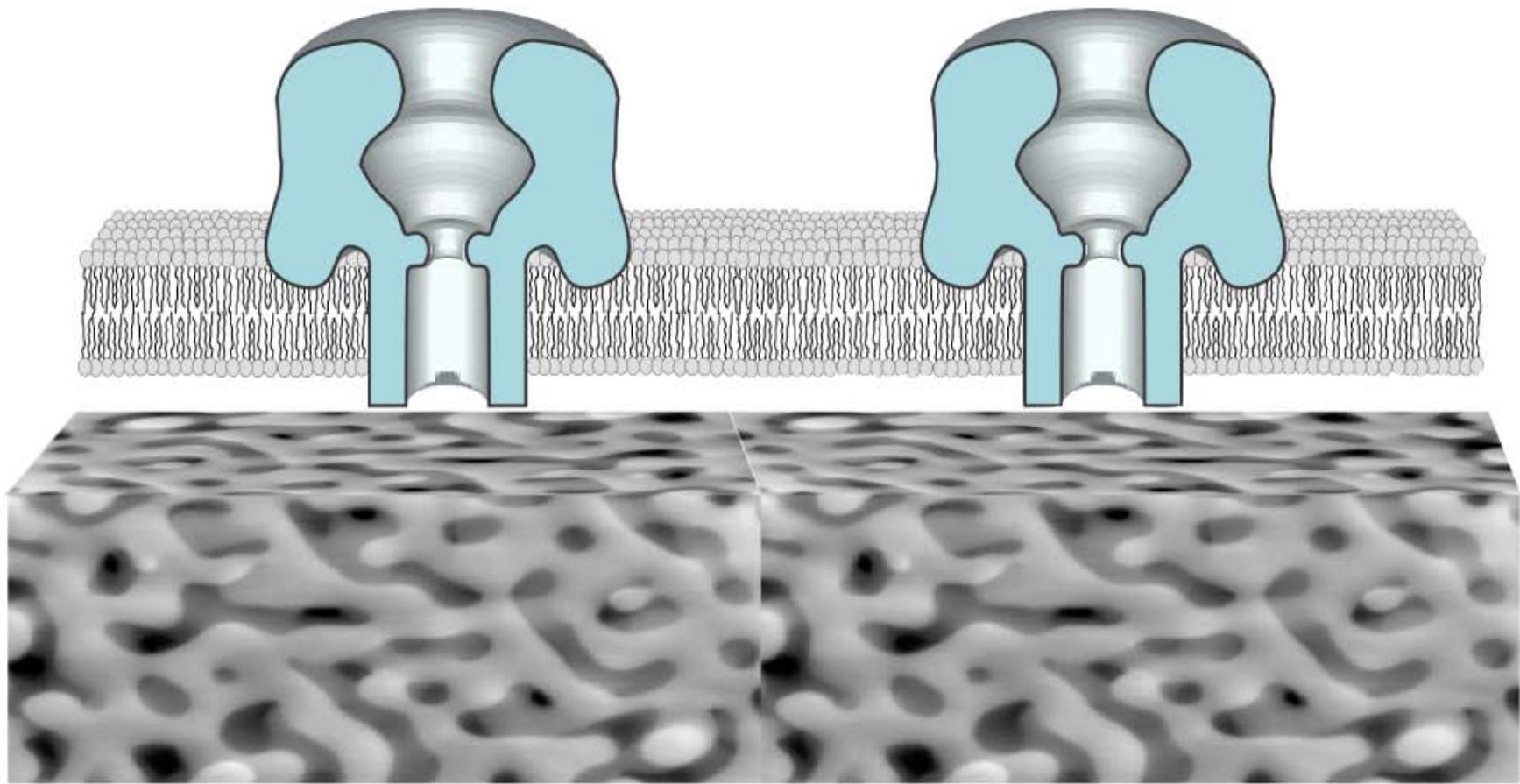


resistance against PLA₂ depends on cover lipid:

- well-ordered DPhyPC largely insensitive
- fluid POPC immediately degraded

Functionalization with PA63: DPhyPC

work in progress toward:



Conclusions

- Neutron scattering is adequate for non-destructive, molecular-scale characterization of molecular interface architectures
- Optimization of tethered membrane structures using EIS and neutron scattering: Membrane properties controlled by backfilling and lipid chemistry
 - Densely grafted SAM insufficiently hydrated in submembrane space
 - Electric field induces water (by introducing membrane defects)
 - “Backfilling” more gentle method to introduce hydration
 - “Rapid solvent exchange” works well on backfilled SAM
 - Great flexibility in choice of lipid for bilayer completion
 - Magnetic contrast variation: Investigation of minute structural details
 - PLA₂: A sensitive amplifier of membrane surface defects

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- Frank Heinrich

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